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# Factor XI and traveling waves: the key to understanding coagulation in hemophilia?

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## "The 'spatial' concept can provide a new point-of-view for understanding the bleeding mechanism and hemophilia treatment strategies."

Hemophilia A, B and C are hereditary bleeding disorders that result from congenital deficiencies or defects of blood coagulation factors VIII (FVIII), IX (FIX) and XI (FXI), respectively. These proteins are part of the so-called intrinsic pathway of the coagulation cascade. FIX is activated by FXIa, and FXI is in turn activated by factor XIIa (FXIIa) that, in its turn, is spontaneously autoactivated when it is in contact with a negatively charged surface. The FXII pathway, however, is not presently considered to be physiologically relevant for normal hemostasis [1]. Currently, extravascular membrane protein tissue factor (TF) is well established as a major factor initiating clotting via the extrinsic pathway. This pathway starts from TF-FVIIa complex, extrinsic tenase, which is formed upon TF exposure at the site of injury. Extrinsic tenase activates FIX and factor X, and factor Xa (FXa) activates prothrombin into thrombin, the main enzyme that fulfills the actual coagulation - that is, formation of a fibrin clot. Thrombin also performs other functions, in particular by rapidly activating factor V and FVIII, elements of positive feedbacks that accelerate FXa and thrombin formation by three to five orders of magnitude. So FVIII and FIX are more or less explicitly involved in both pathways.

By contrast, there was no natural activator for FXI other than FXIIa reported for many years. However, deficiency of FXII is not associated with bleeding. It seemed strange, as a deficiency of FXI (hemophilia C) leads to bleeding, although clinical manifestation is milder compared

with deficiency of FIX and FVIII. It meant that FXI had a function in hemostasis independent of FXII.

A critical progress was achieved two decades ago with the finding that FXI can be activated, albeit slowly, by thrombin [1]. This activation can occur in vivo and enhance FIX generation. However, there is a problem: thrombin cleaves fibrinogen to form fibrin much faster than it activates FXI [1], so FXIa seems to have no considerable effect on clot formation. Thrombin formation is affected more significantly [2,3]. Still, both effects can be observed only in a limited range of experimental conditions: absence of FXI plays a role when there is low or zero TF concentration [2-5]. In these cases, FXI deficiency can lead to delayed clot formation and decreased thrombin generation.

The question of the meaning of thrombin generation actually goes beyond the problem of hemophilia. While various forms of thrombin generation assays have become an established clinical tool, it is also established that fibrin clot is already formed when only 5–10% of thrombin is activated [6]. This raises a major question: why thrombin, produced after the fibrin clot formation, is important?

With respect specifically to hemophilia C, it was proposed that the need for additional thrombin can be explained by the observation that bleeding usually occurs in the tissues with increased fibrinolysis rate [7]. Decreased thrombin generation can reduce protection of the clot to fibrinolysis by the following mechanisms:

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- Structure of fibrin clot depends on thrombin concentration, as lower thrombin concentrations (i.e., in case of hemophilia) form a clot with thicker fibers that are easier to lyze [8];
- Thrombin protects the clot from fibrinolysis by activating TAFI [9];
- Thrombin activates FXIII, which is necessary for stable clot formation.

However, this does not solve all of the problems. As mentioned earlier, thrombin generation depends on FXI in cases of very low TF concentrations (less than 1 pM). The density of TF present at the site of injury, especially during surgery or in severe trauma, however, is very high (e.g., amount of TF on the surface of fibroblasts is ~100,000 molecules/cell) [10]. If we try to relate concentrations and densities, however, another possible solution arises. TF *in vivo* is not dissolved, in contrast to the *in vitro* prothrombin time and thrombin generation assays. It is localized on the damaged tissues so that the clot grows from the surface of the TF-bearing cells to the volume of blood where there is no TF. Therefore, FXI can possibly perform its function in the propagation of coagulation from the activator, because far from the activator there is no another source of FIXa.

The idea of the spatial separation of the reactions of initiation and propagation of coagulation was proposed by Hoffman and Monroe [11,12] as a cell-based model of coagulation and by the authors' group [13,14] as a spatial propagation model. Despite the different experimental systems, it was shown that initiation and propagation of coagulation are regulated by different reactions of the coagulation cascade [12,15].

The basic idea of this approach, as it appears today, is the following: FXa, which is necessary to produce thrombin, is generated by extrinsic tenase on the surface of TF-expressing cells. However, this factor is inhibited very rapidly in plasma, and hence cannot get far from the activator by diffusion (it is protected from the inhibition on the platelet surface, but it needs to get there to get this protection). Without another source of FXa, its generation would be confined to a thin layer of the TF-expressing cells, and no solid 3D fibrin clot would be formed. So, there should be some source of FXa that can diffuse and produce FXa in a new location. This function is performed by FIXa, which has a much lower rate of inactivation by the plasma inhibitors [15] and can diffuse much further from the point of activation. In addition, when FIXa diffusion is not enough, FXIa comes into play. FXI can be activated by thrombin and thus form a source of FIXa that would be independent of TF-initiated coagulation. The clotting process thus could propagate even when TF is no longer available.

Typical coagulation assays simulate the coagulation process close to TF-bearing cells where TF is always present. In these models, the role of FXI is diminished. To emphasize the propagation phase of coagulation, the experimental system should be considered where it is possible to observe clot growth from the TF-coated surface in plasma. In this system, hemophilia A and B were shown to be associated with abnormal clot growth, while initiation by TF was not affected [10].

Recently, we were able to directly measure thrombin distribution in the growing clot *in vitro* [16]. High concentration of thrombin is produced near the activating TF-coated surface. The concentration of thrombin produced is dependent on TF concentration and is similar to the thrombin peak obtained in regular thrombin generation assay. However, there is the second phase of thrombin generation in the spatial system. Clot propagation in space is driven by a moving peak of high thrombin concentration. This peak is fully independent from the TF and is propagating with a constant speed far from the activator. In addition, in these conditions, the only way to sustain thrombin generation independent from TF is activation of FIX by FXIa.

This self-sustaining wave of thrombin is not formed in the absence of factor XI (as well as FVIII), showing that this TF-independent mechanism is required for clot growth. This mechanism of coagulation propagation can spread the coagulation process far enough from the damaged tissue. It can be especially important in cases of large-scale injury such as trauma or surgery, in situations when bleeding is observed in hemophilia C.

This concept is in accordance with the previous results [3,4,9] and can reinforce the 'fibrinolysis' hypothesis. In the absence of FXI, the moving thrombin peak is not formed and the local concentration of thrombin far from the activator at the time of clot formation is much lower [16]. This can lead to an increased fibrinolysis rate for the reasons mentioned above.

Considering coagulation as a spatially heterogeneous process shows that thrombin production near the TF-covered surface and far from it are completely different processes controlled by different reactions of the coagulation cascade. FXI is important in the TF-independent propagation phase of clot formation, not in the initiation process.

This new concept of bleeding in hemophilia can lead to another view on diagnostics and therapy. Specifically, experiments with existing bypassing agents [17] and potential new ones [18] indicate that this should be kept in mind: thrombin produced near activation site is not equivalent to that produced far from it. The 'spatial' concept can provide a new point-of-view for understanding the bleeding mechanism and hemophilia treatment strategies. Additional experiments, in particular in animal models, should be performed to elucidate this in more detail.

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# Factor XI & traveling waves Editorial

### References

- Gailani D, Broze GJ Jr. Factor XI activation in a revised model of blood coagulation. *Science* 253(5022), 909–912 (1991).
- 2 Keularts IM, Zivelin A, Seligsohn U, Hemker HC, Béguin S. The role of factor XI in thrombin generation induced by low concentrations of tissue factor. *Thromb. Haemost.* 85(6), 1060–1065 (2001).
- 3 Cawthern KM, van't Veer C, Lock JB, DiLorenzo ME, Branda RF, Mann KG. Blood coagulation in hemophilia A and hemophilia C. *Blood* 91(12), 4581–4592 (1998).
- 4 Wielders SJ, Béguin S, Hemker HC, Lindhout T. Factor XI-dependent reciprocal thrombin generation consolidates blood coagulation when tissue factor is not available. *Arterioscler. Thromb. Vasc. Biol.* 24(6), 1138–1142 (2004).
- 5 Rugeri L, Quélin F, Chatard B, De Mazancourt P, Negrier C, Dargaud Y. Thrombin generation in patients with factor XI deficiency and clinical bleeding risk. *Haemophilia* 16(5), 771–777 (2010).
- 6 Mann KG, Brummel K, Butenas S. What is all that thrombin for? J. Thromb. Haemost. 1(7), 1504–1514 (2003).

- 7 Smith SB, Gailani D. Update on the physiology and pathology of factor IX activation by factor XIa. *Expert Rev. Hematol.* 1(1), 87–98 (2008).
- 8 Wolberg AS. Thrombin generation and fibrin clot structure. *Blood Rev.* 21(3), 131–142 (2007).
- 9 von dem Borne PA, Meijers JC, Bouma BN. Feedback activation of factor XI by thrombin in plasma results in additional formation of thrombin that protects fibrin clots from fibrinolysis. *Blood* 86(8), 3035–3042 (1995).
- 10 Ovanesov MV, Krasotkina JV, Ul'yanova LI *et al.* Hemophilia A and B are associated with abnormal spatial dynamics of clot growth. *Biochim. Biophys. Acta* 1572(1), 45–57 (2002).
- 11 Hoffman M, Monroe DM, Oliver JA, Roberts HR. Factors IXa and Xa play distinct roles in tissue factor-dependent initiation of coagulation. *Blood* 86(5), 1794–1801 (1995).
- 12 Monroe DM, Hoffman M, Roberts HR. Transmission of a procoagulant signal from tissue factor-bearing cell to platelets. *Blood Coagul. Fibrinolysis* 7(4), 459–464 (1996).
- 13 Ataullakhanov FI, Volkova RI, Guriia GT, Sarbash VI. Spatial aspects of blood

coagulation dynamics. III. Growth of clots in vitro. Biofizika 40(6), 1320–1328 (1995).

- 14 Ataullakhanov FI, Guria GT, Sarbash VI, Volkova RI. Spatiotemporal dynamics of clotting and pattern formation in human blood. *Biochim. Biophys. Acta* 1425(3), 453–468 (1998).
- 15 Panteleev MA, Ovanesov MV, Kireev DA et al. Spatial propagation and localization of blood coagulation are regulated by intrinsic and protein C pathways, respectively. *Biophys. J.* 90(5), 1489–1500 (2006).
- 16 Dashkevich NM, Ovanesov MV, Balandina AN *et al.* Thrombin activity propagates in space during blood coagulation as an excitation wave. *Biophys. J.* 103(10), 2233–2240 (2012).
- 17 Ovanesov MV, Panteleev MA, Sinauridze EI *et al.* Mechanisms of action of recombinant activated factor VII in the context of tissue factor concentration and distribution. *Blood Coagul. Fibrinolysis* 19(8), 743–755 (2008).
- 18 Parunov LA, Fadeeva OA, Balandina AN *et al.* Improvement of spatial fibrin formation by the anti-TFPI aptamer BAX499: changing clot size by targeting extrinsic pathway initiation. *J. Thromb. Haemost.* 9(9), 1825–1834 (2011).